

## Efficiency of Using Different Methionine Sources in Low Methionine Diets of (*Oreochromis niloticus*) Fingerlings

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### SUMMARY

An experiment was conducted to evaluate the use of different crystalline methionine sources to optimize amino acid profiles and its effect on growth performance and feed utilization in feeds of all male tilapia *O. niloticus*. Seven treatments were applied, one basal diet as negative control which contains 3.9 g kg<sup>-1</sup> Methionine (MET) and diets (2-6) were supplemented with two levels of MET dietary (0.15–0.30%) of DL- MET, novel MET1 and MET2 at a constant dietary cysteine level of 3.9 g kg<sup>-1</sup>, respectively. The tested diets formulated to contain 28% crude protein and 18.7 MJ GE, applied in 28 fiberglass tanks, 25 Nile tilapia fingerlings (13.5 ± 0.40g). Growth performance and feed efficiency were improved significantly for tilapia fed on diets supplemented by different MET products compared to non supplemented diet (control). The present study clearly shows that optimizing the amino acid profile by inclusion of a single crystalline amino acid (crystalline MET) does not give the same protein utilization as that of a diet in which the amino acids are mainly protein bound. Diet containing MET1 improved protein utilization than DL- MET or MET2.

**Keywords:** amino acid, efficiency, requirement, growth performance, feed utilization, tilapia

### INTRODUCTION

As fish meal is an expensive component of commercial fish feeds and beyond sustainable limits (Cho and Bureau, 2001), fish nutrition investigations are mainly directed towards reducing feed cost by manipulating the feed formulation. Most published research focused on plant protein sources such as soybean meal (Wilson *et al.*, 2004), cottonseed meal (Lim, 1996) and sunflower meal (Maina *et al.*, 2003). However, plant feedstuffs mostly associate with lower nutrient concentrations, insufficient levels of amino acids (AA) (e.g. lysine and methionine), anti-nutritional factors and poor palatability (Tacon and Jackson, 1985 and Denstadli *et al.*, 2006). Furthermore, methionine is one of the most limiting essential AA in plant protein sources for fish diet formulation (Goff and Gatlin 2004; Alam *et al.* 2005; Zhou *et al.*, 2006). Therefore, it is expected that extra methionine has to be added to plant protein based diets in order to promote optimal growth and health of fish. Thus, to maximize the utilization of plant protein sources, especially those that have low methionine levels, the determination of methionine requirement is important for the development of cost-effective fish feeds. In this context, it is also relevant to investigate the efficiency of methionine supplementation to practical diets that are expected to be deficient in methionine supply. To date, amino acid requirement studies in fish mostly make use of dose-response effects on growth, nutrient deposition and/or several physiological criteria and utilize graded supplementation of the crystalline amino acid under study (Shearer 2000; Wilson 2003 and Kaushik and Seiliez, 2008). Several studies have been conducted to determine the methionine requirement and it was

found to be 0.1, 0.5, 0.53 and 0.75 % of diet according to Ogunjiet *al.* (2005), Kasper *et al.* (2000), Jackson and Capper (1982) and Santiago and Lovell (1988), respectively, in addition to the total sulphur amino acid requirement of juvenile tilapia was 0.85 % of diet as reported by Nguyen and Davis (2008). Generally, high variation was yielded due to the observed requirement concentration in the final diets. Different approaches are one factor of influence. Supplementation technique requires significant dose-response effects coming from several equally spaced increments of the supplemented limiting AA (Cowey, 1992). Several studies indicated the importance of the adequacy of the breakpoint analysis for obtaining reliable data (Robbins *et al.*, 1979 and Were, 1989). In addition, overdosing of individual amino acids may deliver amino acid imbalances (Wilson, 2003 and Helland *et al.*, 2006). To overcome several disadvantages of supplementation technique, the current study concluded quantitative methionine requirements in juvenile tilapia by use of a nonlinear N-utilization model. Making use of nonlinear N-utilization models and adequate mathematical description of protein deposition as a function of the intake of the first limiting dietary amino acid is one of the important approaches mostly applied for modelling of AA requirements in fish (Liebert *et al.*, 2006; Liebert and Benkendorff, 2007a, b and Liebert, 2009). The current study was conducted to measure the dietary efficiency of protein-bound Met or as supplemented AA for further evaluation of Met requirement data in tilapia feeding.

## MATERIAL AND METHODS

Table 1. Composition (g kg<sup>-1</sup>) of Tilapia diets limiting in methionine supply

Ingredients	Diet						
	1	2	3	4	5	6	7
Corn	240	240	240	240	240	240	240
Soybean meal	250	250	250	250	250	250	250
SPC <sup>1</sup>	150	150	150	150	150	150	150
Peas	100	100	100	100	100	100	100
bean	40	40	40	40	40	40	40
Fish oil	45	45	45	45	45	45	45
Vit Min Mix <sup>2</sup>	10	10	10	10	10	10	10
MCP <sup>3</sup>	10	10	10	10	10	10	10
CaCO <sub>3</sub> <sup>4</sup>	7	7	7	7	7	7	7
CMC <sup>5</sup>	20	20	20	20	20	20	20
L-Thr	0.7	0.7	0.7	0.7	0.7	0.7	0.7
DL- Met	-	1.5	3	-	-	-	-
Met- (P1) <sup>6</sup>	-	-	-	1.5	3	-	-
Met- (P2) <sup>7</sup>	-	-	-	-	-	1.5	3
Wheat starch	127.3	125.8	124.3	125.8	124.3	125.8	124.3
Total	1000	1000	1000	1000	1000	1000	1000
<b>Chemical composition</b>							
Crude protein	275	272	277	277	275	277	276
Crude lipid	82.0	69.3	69.5	70.9	68.6	68.2	93
Ash	67.7	67.5	67.0	66.9	67.5	66.6	66.3
Crude fibre	49.3	49.2	52.7	53.0	54.0	52.6	51.8
N-free extract	526	542	533.8	532.2	534.9	535.6	512.9
Gross energy (kJ g <sup>-1</sup> )	18.8	18.2	18.3	18.4	18.2	18.3	19.2
P:E (g MJ <sup>-1</sup> )	14.7	14.9	15.1	15.1	15.1	15.2	14.4
Methionine	3.9	5.5	7.1	5.4	7.0	5.5	7.0
Methionine+cystine	7.8	9.4	1.10	9.4	1.11	9.6	1.11
Lysine	16.1	16.0	16.0	16.1	15.9	16.1	16.0
Threonine	11.5	11.5	12.3	12.2	11.3	11.8	11.2

1- Soybean protein concentrate

2- Vitamin and mineral mix (provided per kg of diet): MnSO<sub>4</sub>, 40 mg; MgO, 10 mg; K<sub>2</sub>SO<sub>4</sub>, 40 mg; ZnCO<sub>3</sub>, 60 mg; KJ, 0.4 mg; CuSO<sub>4</sub>, 12 mg; ferric citrate, 250 mg; Na<sub>2</sub>SeO<sub>3</sub>, 0.24 mg; Co, 0.2 mg; vitamin A, 4000 IU; vitamin B6, 30 mg; vitamin D3, 400 IU; vitamin E, 400 mg; vitamin B12, 80 µg; vitamin B1, 30 mg; vitamin B2, 40 mg; vitamin K3, 12 mg; folic acid, 10 mg; biotin, 3 mg; pantothenic acid, 100 mg; inositol, 50 mg; ascorbic acid, 500 mg.

3- Calcium carbonate

4 – Monocalcium phosphate

5-Carboxyl methyl cellulose

6,7- Evonik Degussa GmbH, RodenbacherChaussee 4, 63457 Hanau, Germany

### 1. Experimental diets:

Seven iso-nitrogenous and iso-caloric diets were formulated from practical ingredients to contain almost 28% crude protein 18.7 MJ GE (Table1).The diets were based on plant protein sources (Corn, Soybean meal, soybean-condensed protein, peas and

bean). One basal diet as negative control which contain 3.9g kg<sup>-1</sup> Methionine and 6 diets were supplemented with two levels of methionine dietary (0.15–0.30%) DL-methionine, novel methionine P1 and methionine P2 (Six diets were prepared by adding DL-Met, or two Met sources consisting of Met dimmers, including either DD-,LL-,DL-,and LD-Met

dimmers (MET1) or mixture of DL- and LD-Met (MET2) at a constant dietary cystine level of  $3.9 \text{ g kg}^{-1}$ , respectively. Crystalline amino acid (L-Therionine) was added to all diets to cover the amino acid requirements of tilapia according to (NRC 2011). Fish oil was added as a major dietary lipid source to make all diets isolipidic. Wheat starch was used as additional source of energy to adjust the energy content of the diets. The vitamin mixture used was a commercial mixture of (Deutsche Vilomix Tiernahrung GmbH) which was added to all experimental diets at a constant level of 1%. The wet mixture was passed through granule machine with 2 mm diameter. The produced pellets were dried at room temperature for three days (approximately 10 % moisture was achieved). The dried pellets were stored in a cool room at  $2^\circ\text{C}$ .

## 2. Experimental conditions:

Experimental fish were stocked in a semi-closed in-door water recirculation system with 28 circular plastic tanks (250-L/tank). Each tank was continuously supplied with a mixture of fresh water (approximately 10%) and biologically filtered fresh water. The photoperiod (12h light: 12h dark) were regulated. The water temperature was recorded daily and other quality parameters including pH, ammonia and  $\text{NO}_2$  were recorded weekly.

## 3. Experimental fish:

Tilapia, *Oreochromis niloticus* were obtained from Department of Animal Science, Division of Animal Breeding and Animal Genetics, Göttingen University, Germany. Fish were acclimated to laboratory conditions for two weeks before being distributed into plastic tank of 250-L water capacity each. Fish with an average of  $13.5 \pm 0.4 \text{ g}$  initial body weight were distributed into 28 circular experimental tanks four replicate groups of 25 fish each. The experiment lasted 56 days. All fish in each tank were weighed every 14 days. During the growth period, each diet was offered to fish groups by hand, 4 meals/day.

## 4. Sample collection:

At the beginning of the experiment, ten fish were analyzed for body composition. Three fish/ tank were sampled at the end of experiment, killed by anesthetic overdose (Ethylene-glycol-monophenyl-ether), autoclaved ( $110^\circ\text{C}$ , 3 h), homogenized with lab mixer and stored at  $-20^\circ\text{C}$  for subsequent chemical analysis. The feed ingredients and diets, which were used in the experimental study, were also chemically analyzed according to the method described by NAUMANN and BASSLER (1976 – 1997) in duplicates. Dry matter determination used an oven at  $110^\circ\text{C}$  (Memmert) until constant weight; crude ash was detected by 4 h ashing at  $600^\circ\text{C}$  in a furnace muffle (Thermicon P; Heraeus Holding). A nitrogen auto-analyzer (FP-2000; Leco) was utilized for crude protein determination using the Dumas-method ( $\text{N} \times 6.25$ ). Ether extract was determined by

extraction with petroleum ether according to the Soxhlet-procedure following HCl-hydrolysis of the feed samples. Gross energy of the diets was calculated due to NRC (1993), based on crude nutrient analyses. Amino acids analyses (except tryptophan) were conducted by ion-exchange chromatography (LC 3000, Biotronic, Eppendorf-Netheler-Hinz GmbH, Hamburg, Germany) following acid hydrolysis with and without an oxidation step for determination of sulphur-containing Amino acids.

## 5. Statistical analysis:

All data of growth performance and feed utilization were analyzed by one-way analysis of variance (ANOVA) using the general linear models procedure of statistical analysis system (SPSS) version 17, (2009). Duncan's multiple range test (Duncan, 1955) was used to resolve differences among treatment means at 5% significant level using the following model.  $Y_{ij} = \mu + T_i + E_{ij}$

Where

$\mu$  = over all mean.

$Y_{ij}$  = the observation of the individual from T treatment

$T_i$  = the fixed effect of T diet.

$E_{ij}$  = the experimental random error associated with individual J.

The model for assessing the dietary AA-efficiency

According to equations 1 and 2, the applied model basically describes body nitrogen retention (NR) depending on nitrogen intake (NI) and dietary protein quality (b):

$$(1) \text{NR} = \text{NR}_{\max} \text{T} (1 - e^{-b \cdot \text{NI}})$$

$$(2) \text{ND} = \text{NR}_{\max} \text{T} (1 - e^{-b \cdot \text{NI}}) - \text{NMR}$$

Where:

$$\text{NR} = \text{daily N-retention (ND + NMR)} \\ [\text{mg}/\text{BW}_{\text{kg}}^{0.67}]$$

$$\text{ND} = \text{daily N-deposition}$$

$$[\text{mg}/\text{BW}_{\text{kg}}^{0.67}]$$

$\text{NMR} = \text{daily N-maintenance requirement}$

$$[\text{mg}/\text{BW}_{\text{kg}}^{0.67}]$$

intake

$$[\text{mg} / \text{BW}_{\text{kg}}^{0.67}]$$

$\text{NR}_{\max} \text{T} = \text{Theoretical maximum for daily N-retention}$

$$[\text{mg} / \text{BW}_{\text{kg}}^{0.67}]$$

$$\text{ND}_{\max} \text{T} = \text{NR}_{\max} \text{T} - \text{NMR}$$

b = slope of function (1) and (2), respectively

(Indicating the dietary protein quality independent on the N-intake)

e = basic number of natural logarithm

(ln)

Estimation procedure of the model parameters  $\text{NR}_{\max} \text{T}$ ,  $\text{ND}_{\max} \text{T}$ , and NMR was reported for all male juvenile *O. niloticus* (Liebert et al., 2006) and also utilized for the current investigations ( $\text{NMR} = 70 \text{ mg} / \text{BW}_{\text{kg}}^{0.67} / \text{day}$ ;  $\text{NR}_{\max} \text{T} = 388 \text{ mg} / \text{BW}_{\text{kg}}^{0.67} / \text{day}$ ;  $\text{ND}_{\max} \text{T} = 318 \text{ mg} / \text{BW}_{\text{kg}}^{0.67} / \text{day}$ ). Following

logarithmization and transformation of equation (1), further model parameters are obtained:

$$(3) \quad b = [\ln NR_{\max}T - \ln (NR_{\max}T - NR)] : NI$$

According to equation (3) the model parameter "b" is determined, indicating the dietary protein quality. Equation (4) calculates the required NI for a defined NR depending on feed protein quality (b):

$$(4) \quad NI = [\ln NR_{\max}T - \ln (NR_{\max}T - NR)] : b$$

The fundamental relationship between intake of the limiting amino acid (LAAI) and NR provides equation (5):

$$(5) \quad LAAI = [\ln NR_{\max}T - \ln (NR_{\max}T - NR)] : [16 \cdot bc^{-1}]$$

Where

LAAI = daily intake of the LAA  
 $[mg/BW_{kg}^{0.67}]$   
 (Indicating the requirement dependent on NR and  $bc^{-1}$ )  
 $bc^{-1}$  = Slope between LAA-concentration (c) and feed protein quality (b)  
 (The slope is indicating the efficiency of dietary LAA-utilization)

Consequently, LAAI depends on the aimed performance (NR) and the observed efficiency of utilization of the limiting AA ( $bc^{-1}$ ). Efficiency in context with the exponential model integrates digestion, absorption and post-absorptive utilisation of the LAA (Samadi and Liebert 2006, 2007), more details about model application for fish are reported elsewhere (Liebert and Benkendorff 2007a, b; Liebert 2009).

## RESULTS

The survival rate of Nile tilapia after 8wk of feeding experimental diets was 100%. All the water quality parameters were within the acceptable range for Nile tilapia dissolved oxygen,  $6.56 \pm 0.27$  mg/L; water temperature,  $28.3 \pm 0.4$  °C; ammonia,  $0.070 \pm 0.062$  mg/L; nitrite-N,  $0.041 \pm 0.028$  mg/L; and pH,  $8.0 \pm 0.2$ . These values were within optimum ranges for normal growth and health of juvenile Nile tilapia (Watanabe *et al.*, 1993 and El-Shafaiet *et al.*, 2004). Average of initial body weight of Nile tilapia fingerlings fed the experimental diets at the start did not differ, indicating that groups were homogenous. Growth data from experiment study are shown in Table (2). At the end of the experimental period (56days), the group of fish fed on the supplemented diets containing methionine products (DL- MIT, DD-, LL-, DL-, and LD-Met dimers (MET1) or mixture of DL- and LD- methionine (MET2) grew as well or better than the group of fish fed the control diet. Diets 3, 5 and 7 had significantly ( $P < 0.05$ ) higher final body weight than the rest of the experimental groups. However, the lowest final body weight (29.9g) was achieved in the group of fish fed on the control diet. Analysis of variance for weight gain (WG) and (SGR) followed the same trend as in final body

weight. However, the increasing of methionine level up to  $7g \text{ kg}^{-1}$  (dry diet) in diets 3, 5 and 7 had significantly ( $P < 0.05$ ) higher SGR than the rest of experimental groups compared to control diet Table 2. There was a significant ( $P < 0.05$ ) improvement in (FCR) and (PER) when the feeds were supplemented with DL-MIT or DD-, LL-, DL-, and LD-Met dimmers (MET1) or mixture of DL- and LD- methionine as compared with the non-supplemented of protein diets. Protein retention, measured as protein retention efficiency (PRE), was better in diets supplemented by different products of methionine compared to the control or diet 2. The average of feed conversion ratio (FCR) in fish groups fed on diets 3 and 5 followed by groups of fish fed on diets 7 were significantly ( $P < 0.05$ ) improved in comparison with the other groups and better than the control diet. The FCR was found to be 1.91 (control diet), 1.79, 1.61, 1.73, 1.68, 1.75 and 1.66, respectively. These results showed that the best ( $P < 0.05$ ) FCR values were obtained by groups of fish fed on diet 3, 5 followed by diet 7. The best FCR values observed with diets supplemented with by methionine product at level of 0.30% (DL-MIT or DD-, LL-, DL-, and LD-Met dimmers (MET1) or mixture of DL- and LD- methionine (MET2) suggesting that addition of methionine product improved feed utilization. The PER found to be 2.47, 2.47 and 2.42 respectively. These results indicate that the best ( $P < 0.05$ ) PER values were obtained by groups of fish fed on diet 3, 5 followed by diets 7. The efficiency of different methionine product were confirmed by results of protein quality (PER) evaluation as yielded from application of an exponential N-utilization model, eliminating the influence of varying feed intake on response of protein deposition (Table 4). The results of PER in fish group fed on diet 5, followed by groups fed on diets 3 and 7 were significantly ( $P < 0.05$ ) improved in comparison with the other groups and better than the control diet. This indicates that the best results of PER (37.5%) obtained by group of fish fed on diet 5 which was supplemented by DL-, DD-, LL-, DL-, and LD-Met dimmers at level of 0.30% ( $7g \text{ kg}^{-1}$  MET1). Body composition data of the fish fed diets with various levels of methionine products in the diet are presented in Table 3. Data clearly indicate that various levels of dietary Methionine had significant ( $P < 0.05$ ) effect on the body composition of *O. niloticus*. Dry matter content remained almost similar among the groups of fish fed on experimental diets except group of fish fed on diet 7 which showed low dry matter content. Ash content was decreased significantly ( $P < 0.05$ ) among the groups receiving diets up to  $7g \text{ kg}^{-1}$  Methionine in the diet. Body protein tended to increase significantly ( $P < 0.05$ ) with increasing dietary methionine concentrations up to  $7g \text{ kg}^{-1}$  of the diet (Diet 7). However, significant ( $P < 0.05$ ) in body lipid content was noted for those fed diet with  $7g \text{ kg}^{-1}$  Methionine (Diet 7) as compared with control diet.

**Table 2. Growth data of tilapia fingerling fed experimental diets limiting in methionine supply**

Parameters	Diet						
	1	2	3	4	5	6	7
Initial weight (g)	13.48	13.36	13.46	13.5	13.31	13.45	13.36
Final weight (g)	29.9±1.9 <sup>c</sup>	30.8±2.2 <sup>ab</sup>	33.13±2.2 <sup>ab</sup>	31.7±1.8 <sup>ab</sup>	32.3±1.4 <sup>ab</sup>	31.6±1.4 <sup>ab</sup>	32.3±1.7 <sup>a</sup>
WG <sup>1</sup> (%)	122±14 <sup>c</sup>	130±16 <sup>ab</sup>	146±15 <sup>a</sup>	134±13 <sup>ab</sup>	143±11 <sup>ab</sup>	134±11 <sup>ab</sup>	142±13 <sup>ab</sup>
SGR <sup>2</sup> (%/d)	1.42±0.1 <sup>c</sup>	1.49±0.1 <sup>b</sup>	1.60±0.2 <sup>a</sup>	1.52±0.1 <sup>b</sup>	1.58±0.9 <sup>a</sup>	1.52±0.1 <sup>b</sup>	1.58±0.8 <sup>a</sup>
Feed intake (g)	787±69 <sup>a</sup>	798±90 <sup>a</sup>	791±88 <sup>a</sup>	786±53 <sup>a</sup>	775±33 <sup>a</sup>	793±15 <sup>a</sup>	783±42 <sup>a</sup>
FCR <sup>3</sup> (g/g)	1.91±0.1 <sup>c</sup>	1.79±0.1 <sup>ab</sup>	1.61±0.0 <sup>a</sup>	1.73±0.1 <sup>a</sup>	1.68±0.9 <sup>a</sup>	1.75±0.1 <sup>ab</sup>	1.66±0.1 <sup>a</sup>
PER <sup>4</sup> (g/g)	2.09±0.1 <sup>c</sup>	2.21±0.1 <sup>bc</sup>	2.47±0.1 <sup>a</sup>	2.30±0.1 <sup>b</sup>	2.47±0.1 <sup>a</sup>	2.27±0.1 <sup>b</sup>	2.42±0.12 <sup>a</sup>
PRE <sup>5</sup> (%)	30.8±1.9 <sup>c</sup>	33.1±1.0 <sup>bc</sup>	36.8±0.8 <sup>a</sup>	34.5±1.1 <sup>ab</sup>	37.5±0.9 <sup>a</sup>	35.5±1.0 <sup>ab</sup>	35.7±0.9 <sup>ab</sup>

Values in the same row with a common superscript are not significantly different (P<0.05).

<sup>1</sup>) Weight gain = 100 × (final body weight- initial body weight)/initial body weight.

<sup>2</sup>) Specific growth rate= (Ln. Final body weight- Ln. Initial body weight) x 100/ experimental period (days)

<sup>3</sup>) Feed conversion ratio = g dry feed consumed/g wet weight gain.

<sup>4</sup>) Protein efficiency ratio = fish wet weight gain/protein intake.

<sup>5</sup>) Protein retention efficiency=100× (final body protein – initial body protein)/total protein fed.

**Table 3. Whole body composition of fingerling tilapia (g kg<sup>-1</sup> dry matter)**

Diet	Initial	1	2	3	4	5	6	7
Dry matter	174.2	220±0.12 <sup>b</sup>	222±0.34 <sup>b</sup>	220±0.76 <sup>b</sup>	223±0.16 <sup>b</sup>	222±0.50 <sup>b</sup>	229±0.44 <sup>a</sup>	215±0.57 <sup>c</sup>
Crude protein	688.6	611±0.70 <sup>c</sup>	14±0.70 <sup>c</sup>	620±0.70 <sup>b</sup>	612±1.41 <sup>c</sup>	623±2.12 <sup>b</sup>	613±1.41 <sup>c</sup>	630±2.8 <sup>a</sup>
Crude lipid	109.6	257±0.71 <sup>a</sup>	230±0.78 <sup>d</sup>	236±0.75 <sup>b</sup>	239±0.9 <sup>b</sup>	229±0.6 <sup>c</sup>	233±0.5 <sup>c</sup>	227±0.8 <sup>e</sup>
Ash	201.8	132±1.5 <sup>d</sup>	156±0.9 <sup>a</sup>	144±0.0 <sup>c</sup>	149±1.6 <sup>b</sup>	148±0.9 <sup>b</sup>	154±1.5 <sup>a</sup>	143±1.1 <sup>c</sup>

Values in the same row with a common superscript are not significantly different (P<0.05).

**Table 4. Model parameters data of tilapia fingerling fed experimental diets limiting in methionine supply**

Model parameters	Diet						
	1	2	3	4	5	6	7
Daily N-intake <sup>1</sup>	289±19 <sup>a</sup>	285±24 <sup>a</sup>	280±23 <sup>a</sup>	283±15 <sup>a</sup>	275±86 <sup>a</sup>	285±4 <sup>a</sup>	279±10 <sup>a</sup>
Daily N-retention <sup>2</sup>	160±7 <sup>b</sup>	166±8 <sup>ab</sup>	172±7 <sup>a</sup>	168±7 <sup>ab</sup>	171±5 <sup>a</sup>	172±5 <sup>a</sup>	170±6 <sup>a</sup>
Protein quality b (*10 <sup>6</sup> )	1845±96 <sup>c</sup>	1959±15 <sup>bc</sup>	2120±98 <sup>ab</sup>	2014±117 <sup>ab</sup>	2159±97 <sup>a</sup>	2062±102 <sup>ab</sup>	2073±89 <sup>ab</sup>
Met efficiency bc-1 (*10 <sup>6</sup> )	1205±62 <sup>c</sup>	913±54 <sup>b</sup>	763±35 <sup>a</sup>	944±55 <sup>b</sup>	774±35 <sup>a</sup>	963±47 <sup>b</sup>	746±32 <sup>a</sup>
Met content cMet (g/16gN)	1.53	2.14	2.77	2.13	2.78	2.14	2.77

Values in the same row with a common superscript are not significantly different (P<0.05)

<sup>1</sup>NI(mg / BWkg<sup>0.67</sup>), <sup>2</sup>NR(mg / BWkg<sup>0.67</sup>)

## DISCUSSION

The utilization of crystalline amino acids in practical diets of fish depends on the species ability to absorb and use such amino acids for protein synthesis as well as for other physiological functions. In tilapia, varying results have been obtained with crystalline amino acid supplementation. Growth performance of *Oreochromis niloticus* was improved when a 35%

protein diet formulated from casein was supplemented with either arginine and lysine or tryptophan and methionine (Teshima *et al.*, 1986). Taconet *et al.* (1983) reported that growth of Nile tilapia was improved to a level comparable to that obtained from a fish meal-based diet when 0.8% D, L-methionine was supplemented to a diet in which 75% of brown fish meal was replaced by soybean meal. Further studies with tilapia, which were recently

reported, confirm such findings (Jintasataporn *et al.*, 2010 and Zhu *et al.*, 2010). These results are in agreement with present study, which reported that weight gain of juvenile Nile tilapia was increased significantly as Methionine levels increased by supplementation of crystalline Methionine, indicating that juvenile Nile tilapia could use crystalline Methionine efficiently when Methionine levels in practical diets were deficient. However, in other fish species, the supplementation of essential amino acid (EAA) to a low-protein diet reported in channel catfish (Li and Robinson, 1998) and rainbow trout (Yamamoto *et al.*, 2005), showed that the supplementation of EAA in an insufficient protein diet improved the feed efficiency but did not enhance fish growth. While, (Williams *et al.*, 2001) with Asian sea bass, demonstrated that the crystalline-EAA supplementation to an EAA deficient diet increased fish growth and the response was more pronounced for the low protein diet. Yang *et al.*, (2010) demonstrated significant improvement of growth and feed utilization of grass carp can be achieved by L-Lysine sulphate and MHA-Ca supplementation. WG and PR of the tested group improved 11.4% and 8.3%, respectively, while FCR decreased 9.6% in comparison with control diet. In contrary with these findings, growth and feed efficiency of hybrid tilapia, *O. niloticus* x *Oreochromis aureus*, were not improved when either methionine and lysine or threonine were added to a 24% protein diet containing soybean meal as the main protein source (Liouet *et al.*, 1986). Same trend was reported by (Nguyen 2007 and Nguyen, 2009) which demonstrated that supplementation of methionine to practical diets, did not improve growth, survival and, feed conversion ratio of juvenile tilapia, *Oreochromis* spp. This may be because methionine levels in non-supplemented diets have met the requirement. It is obvious that the methionine level of the control diet are further reduced from their requirements which mainly explained the significant improvement of PER and PRE, as well as the apparent decrease of FCR in the tested group compared with the control group (Table 2). The improved of PRE in this study is in agreement with (Sveier *et al.*, 2001) who reported that supplementation of D-methionine showed better protein utilization than L-methionine when measured as PPV in diet of Atlantic salmon (*Salmon salar* L.) . Same trend was reported by (Yamamoto *et al.*, 2005) who suggested that the EAA supplementation to low-protein diets in rainbow trout improves dietary amino acid utilization and results in an increase of body protein deposition, as similarly noted in gilthead sea bream (Peres and Oliva-Teles, 2009). The body composition of Nile Tilapia is another important aspect, which needed to pay attention to in this experiment (Table 3). The significantly lower whole body lipid and the higher protein of tested fish compared with the control fish indicated a better health condition and higher economic value of tilapia

were obtained by feeding supplemental EAA. However, such result of body lipid was contradictory with an earlier report of grass carp (Wang *et al.*, 2005) and studies of redseabream and freshwater catfish (Chatzifotis *et al.*, 1996 and Tantikitti and Chimsung, 2001) which had higher body lipid content occurred in the lysine supplemented groups. The authors suggest that the amino acids in excess from disproportionate absorption or the part of lysine beyond optimal level would be no longer used for transformation into protein or protein synthesis, but was consumed in catabolism and provided the carbon skeletons for lipid synthesis and deposited as tissue fat, mainly at mesentery. On contrary, opposite trend of whole body lipid content in the present study may be attributed to the insufficient supplementation of methionine, which makes such metabolic pathway impossible. This can be confirmed by the markedly increased whole body protein level of the tested fish, as also noted by Peres and Oliva-Teles (2009) in gilthead sea bream and Yang *et al.* (2010) with Grass Carp (*Ctenopharyngodon idella*).

## CONCLUSION

Supplementation of either Met source yielded dose dependent responses, suggesting general evidence for supplemental amino acids being utilized for growth and protein deposition in fish. Making use of the nitrogen utilization model, the observed efficiency of total Met in diets supplemented with different Met-sources was lower when compared with the non-supplemented basal diet. This observation provides some advice, that utilization of added Met was somewhat lower than utilization of the protein-bound AA. These observations need more investigations in context with the specific features of protein metabolism in fish, comparing DL-Met with the new Met-sources under study (MET1, MET2), an advantage was established of growth and protein utilization as PRE of Nile tilapia can be achieved by the new Met-sources in this investigations.

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## كفاءة استخدام مصادر الميثيونين المختلفة في علائق أصبغيات البلطي النيلي منخفضة الميثيونين

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أجريت تجربة لتقييم استخدام مصادر مختلفة للحامض الأميني الميثيونين في تحسين شكل وتركيب الأحماض الأمينية وتأثيرها على أداء النمو والاستفادة الغذائية في علائق ذكور البلطي النيلي وحيد الجنس. تم تطبيق سبع معاملات، عليقة غذائية أساسية تمثل الكنترول والتي تحتوي على ٣,٩ جم/كجم<sup>١</sup> ميثيونين ، ٢-٦ علائق غذائية تحتوي على مستويين من الميثيونين كمكمل غذائي (٠.١٥ - ٠.٣٠%) من الميثيونين التقليدي، وصور الحامض الأميني ميثيونين الحديثة ومستوي ثابت (٣,٩ جم/كجم) من الحامض الأميني السستينكمكمل غذائي على التوالي. أحتوت العلائق الغذائية المختبرة على ٢٨% من البروتين الخام وطاقة كلية ١٨,٧ ميجاجول. تم تطبيق العلائق التجريبية في ٢٨ حوض من الفيرجلاس خزنت الأسماك بمعدل ٢٥ أصبغية من أسماك البلطي النيلي بمتوسط وزن من ١٣,٥ ± ٠,٤٠ جم عشوائياً في كل حوض. استمرت التجربة لمدة ٥٦ يوم. عموماً، تحسن أداء النمو من حيث الزيادة في الوزن ومعدل النمو النوعي معنوياً في أسماك البلطي المغذاة على العلائق الغذائية المحتوية علي صور مختلفة من الحمض الأميني الميثيونين مقارنة بالعليقة الكنترول. أظهرت نتائج معدل التحويل الغذائي والكفاءة النسبية للبروتين اتجاهات مماثلة لنتائج معدل النمو النسبي في العلائق المحتوية على مكملات الميثيونين الغذائية، ولكن ليس مع العليقة الكنترول. تم قياس كفاءة أحتجاز البروتين وأظهرت النتائج تحسن معدل كفاءة أحتجاز البروتين في العلائق الغذائية المحتوية علي صور مختلفة من الحمض الأميني الميثيونين مقارنة بالعليقة الكنترول. أو العليقة رقم ٢. وتم تأكيد هذه الملاحظات في نتائج تقييم نوعية وجودة البروتين باستخدام نموذج الاستفادة المتزايدة من النيتروجين، بعد تجنب تأثير معدل التغذية علي مقدار الاستجابة لترسيب البروتين. تبين هذه الدراسة بوضوح أن تحسين مجموع الأحماض الأمينية بإدراج الحمض أميني (ميثيونين) لا يعطي نفس الاستفادة من البروتين كمشيلة في العليقة الغذائية المحتوية على أحماض أمينية مرتبطة ببروتين مواد العلف. تحسنت الاستفادة الغذائية من البروتين في العلائق المحتوية على صور الحديثة من الحامض الأميني الميثيونين من النوع الأول عن الميثيونين التقليدي و الصورة الثانية من الحامض الأميني الميثيونين من النوع الثاني.